INFLUENCE OF pH ON VAGINAL DISCHARGES*

BY

LESTER COHEN

Department of Venereology, Cardiff Royal Infirmary

The fluid content of the vagina is derived solely from:

- (1) Mucus secretions of the cervical columnar cells.
- (2) Transudation through the vaginal walls.
- (3) Vulval secretions from sebaceous and sweat glands.
- (4) Bartholin's glands, which are said to produce an "acid mucus secretion".

Vaginal acidity is thought to depend on the presence and amount of lactic acid formed by the action of Döderlein's bacilli on the glycogen of the epithelial cells lining the vagina.

Most pathogenic bacteria have a fairly restricted pH range and grow best at a pH of about 7.5. This may be a reflection of the fact that mammalian tissue and blood have a pH of this order. For example, the pneumococcus has an optimal pH of 7.8 and a growth range of 7.3 to 8.3. Commensal and saprophytic bacteria have a much wider growth range. E. coli has an optimum pH of 6.5 but a growth range of 4.4 to 7.8. Yeasts and fungi generally have an optimum acid pH and may grow at a pH of 2 and lower (Stephenson, 1949). Many of the published data on the pH effects on growth are vitiated because they take into account only the initial pH of the reaction, and also because of inaccurate readings due to inadequate or poorlyhandled equipment. The pH changes so rapidly that the final pH of a reaction may be far removed from that at the start especially where fermentable carbohydrates are present. The reaction at which an organism is grown profoundly affects its enzyme make up; thus, although E. coli will grow at pH 4.5 to 9 (Gale and Epps, 1942), its optimal growth is strictly governed by the pH of optimal activity of the enzymes concerned with nitrogen metabolism (Gale, 1940). Thus, measurement of oxidation reduction potential would be a more accurate reflection of the state of affairs than would pH.

For practical purposes in clinical studies, accurately determined pH values taken under constant conditions have been taken as an acceptable standard of measurement in relation to bacterial, fungal, and protozool activity.

Average vaginal pH values quoted in standard text books

Newborn	5·7
Children	6 to 8
Puberty	4.0
Pregnancy	4.0
Menopause	7.0
Mean pH in the	5.5, falling during ovulation
child-bearing age	

Optimal growth pH of some common organisms

<i>m</i> · 1	4.9 to 7.5 (text-books) 3.6 to 4.7 (Feo, 1956) 5.8 to 6.4 (Lumsden,
Trichomonas vaginalis	₹ 3.6 to 4.7 (Feo, 1956)
	5.8 to 6.4 (Lumsden,
	Robertson, and
	McNeillage, 1966)
Candida albicans	5·4
N. gonorrhoeae	7.5
B. proteus	7.4
Streptococci	7.4
Diphtheroids	7.2

Equipment

A standard model 23A reading pH meter supplied by E.I.L. was used for these studies. Continuous indication is given of the pH value of the liquid under test so that rapidly changing conditions may be followed without difficulty. The machine is fundamentally an ultrastable d.c. amplifier having an exceptionally high input resistance. The output is graduated directly in pH units from 0 to 14, and it is possible with this machine to read accurately to 0.05 pH. The basic stability of the machine is such that zero drift under normal operating conditions

^{*}Received for publication July 24, 1968.

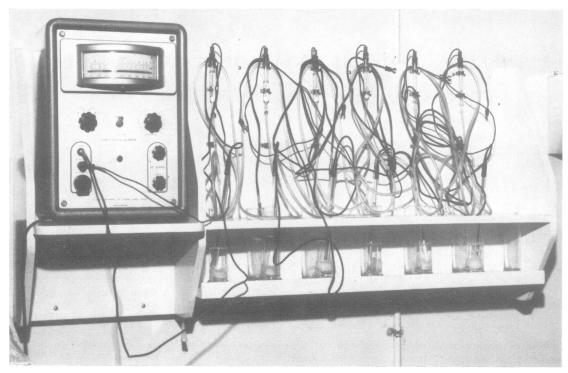


Fig. 1.—Equipment wall-mounted for clinical use.

will not exceed 0.02 pH for periods of 24 hours or more (Fig. 1). All supply voltages are stabilized within the instrument so that its readings remain substantially unaffected by variations in the power line voltage. For use within the vagina the electrodes were designed so that they consisted of a cylinder of silverized glass 23 cm. long and 0.5 cm. in diameter, the contact end of zintered glass being 0.5 cm. in diameter. The calomel reference electrode containing saturated potassium chloride solution was connected by inert tubing to a hollow glass tube 25 cm. long and 0.2 cm. in diameter with a calomel plug at its distal end. The silverized glass electrode had platinum wire running through its length should measurement of oxidation reduction potential be required (Fig. 2, opposite).

This model incorporates a fully automatic temperature compensating mechanism. However, as the temperature of the buffer solution used was prepared and kept at body temperature in a thermostat-controlled jacket and all readings with the machine took place under constant temperature conditions in a draught free centrally heated room at 73°F., there was no need to use the isopotential temperature control system, and a dummy plug resistance supplied with the set was inserted into the machine as the thermometer was not in use.

The electrodes were prepared for use and stood at all times when not in use in sterile distilled water. Two buffer solutions of known pH 4 and 9 were made up

using buffer tablets supplied by Burroughs Wellcome and Co. and closely observing the following points:

- (1) The distilled water was freed of carbon dioxide by boiling.
- (2) The solution was accurately measured to 50 ml. on making up.
- (3) Contamination of the solution with potassium chloride was scrupulously avoided as neutral salts would affect the pH.

With the selector switch set at pH 0 to 14. the electrodes were immersed in the buffer solution of known pH, the set buffer controls were adjusted until the machine read the correct pH value of the buffer solution on the control scale. For initial settings, the makers of the equipment suggest that the electrodes be checked against buffered solutions of known pH 4 and 9 before use, thoroughly rinsing the electrodes in distilled water between measurements.

Method

In clinical use, the pH meter with six sets of electrodes were mounted on a specially designed wall fixture at the right-hand side of the foot of the couch. Each set of electrodes was kept permanently immersed in 100 ml. cylinders containing sterile distilled water with cotton wool pads at the bottom of the jars to prevent damage to the zintered glass. Each cylinder containing distilled water was duplicated by another containing 1 in 1,000

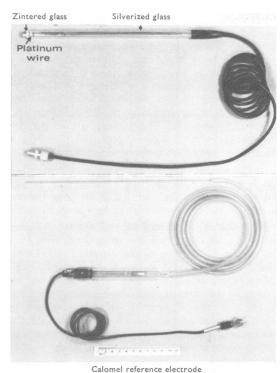


Fig. 2.—Electrodes (detail).

aqueous solution of hibitane. It had previously been found that sterilization of the electrodes in 1 in 1,000 hibitane was the only practical way of avoiding cross-infection and of ensuring that the electrodes were sterile without damaging them. After use, the electrodes were cleaned of contaminating debris and immersed in the hibitane solution for at least 30 minutes. Subsequent incubating of culture media failed to show growth of any bacteria, fungi, or protozoon. Both the distilled water and the hibitane solution were changed twice weekly.

With the patient in the lithotomy position, an unlubricated Cusco speculum was passed, and the cervix displayed as quickly as possible. The balanced electrodes were inserted into the vagina, taking great care to see that they did not touch anything other than the epithelium at the back of the cervix on which the ends were placed in contact. The pH was read directly and the electrodes replaced at once into the hibitane solution after removing debris with gauze. After not less than 30 minutes the electrodes were replaced in buffer solutions for further balancing. With six sets of electrodes in constant use, we have found that there is no danger of using a contaminated electrode. It has been my practice to balance the electrodes every time before use in buffer solutions of pH 4 and 9, thus hoping to ensure absolute accuracy of readings.

At this stage the factors affecting the degree of ionization of a solution must be briefly considered as these will influence the accuracy of the pH readings.

The readings depend on:

- (1) Concentration of the liquid under test;
- (2) Volume of liquid available for analysis;
- (3) Carbon dioxide tension at the time of measurement;
- (4) Temperature;
- (5) Speed of operation.

For practical purposes, the concentration and the volume of the "liquid" under test were constant. There was always sufficient volume of liquid under test conditions for analysis with such a machine. The carbon dioxide tension in the posterior fornix will be constant for each reading provided that the whole operation is carried out speedily. The average time taken to register the reading was in fact 10 sec. and the response time of the machine was 3 sec. There is no variation in temperature from patient to patient that would cause any significant difference in pH. Provided all the above factors are controlled, the pH readings should be accurate with such equipment to 0·1.

Clinical Material

The conditions diagnosed in 200 women (average age 23 years) are shown in Table I. There were 44 cases of gonorrhoea, 58 of infection with *C. albicans*, 42 of vaginitis due only to infestation with *T. vaginalis*, and 56 with essentially normal findings. If those with *T. vaginalis* infestation associated with other venereal infection are added, there were 85 women with trichomonal infestation. In this series

Table I
PREGNANCY AND MARITAL STATUS RELATED TO VAGINAL INFECTION

Diagnosis	No.	Combined Condition	No.	D	Unmarried	
				Pregnant -	No.	Per cent.
Gonorrhoea	44	+ T. vaginalis	30	1	28	66
Non-venereal	56			3	28	50
C. albicans	58	+ T. vaginalis	13	20	12	20
T. vaginalis	42			1	31	80
Total	200		43	25 (12·5 per cent.)	99	50

twenty women with candidiasis were pregnant, all of them married.

Of the total number examined, 50 per cent. were pregnant.

Findings

The distribution of vaginal pH in the whole series of 200 women is shown in Fig. 3, and these values are related to the diagnosis and average age in Table II.

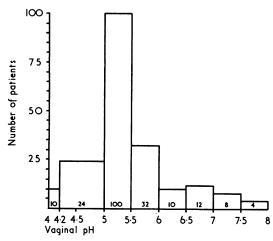


Fig. 3.—Distribution of vaginal pH in a series of 200 women.

Table II
VAGINAL pH VALUES RELATED TO DIAGNOSIS

Discourie.	\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \	p			
Diagnosis	No.	Mean	Range	Average Age (yrs)	
Gonorrhoea	44	5.2	4·2—7·5	22	
Non-venereal	56	5.7	4.0-8.0	21	
C. albicans	58	4.8	4.0—5.5	26	
T. vaginalis	42	5.2	4.0—7.0	30	

The average vaginal pH of the women suffering from gonorrhoea was 5.2 (range 4.2 to 7.5).

This figure is nowhere near the optimal pH of growth of the organism which is 7.5. The average pH of those suffering from vaginal candidiasis is nearer the accepted mark, being 4.8 (range 4.0 to 5.5), the lowest average value of this series. In this group infestation with T. vaginalis was commoner in the slightly older women.

The highest recorded pH in this series was 8 and the lowest 4.0. In the absence of blood in the vagina, the average variation in pH in any one patient did

not exceed 0·2, over a period of 3 to 6 months at any time of the menstrual cycle and irrespective of infection, treatment, type of contraceptive used, type of underclothing worn, and nature of sanitary protection used. The average number of measurements on each patient was 7 (range 4 to 11). The average pH of the 25 pregnant women was 5·5 (range 8 to 4·1), which is far higher than the accepted figure of 4. The average pH of the ten menopausal women was 5·2 (range 4·2 to 5·5), a result far removed from the accepted figure of 7.

In 156 patients the pH lay in the region of 4.2 to 6.0 It was found, as previously stated, that the pH did not vary by more than 0.2 in any one patient during the time she was kept under observation.

A typical cyclic pattern for the vaginal pH does not correspond with that described by Rakoff, Feo, and Goldstein (1944) or by Hunter and Long (1958), who found a pH of 7 at the time of vaginal flow, falling promptly to 5 and tending to remain at this level until the onset of the following menstrual flow. In the presence of even the slightest amount of blood, in some cases even microscopic amounts, the pH reading is always in the region of 7. This is of course never a true reflection of the actual vaginal epithelial pH.

The average pH values in the different age groups (Table III) in this series did not vary by more than 0.9. No children under the age of 10 years were available in this series and no women over the age of 50 years in statistically significant numbers. However, there was nothing in these figures to suggest that in any given person there might be a variation of vaginal pH as wide as from 4 to 8 from birth to menopause including pregnancy. There is no evidence to support the accepted vaginal pH as being 4 at puberty, 4 during pregnancy, and 8 at the menopause.

TABLE III
VAGINAL pH VALUES RELATED TO AGE

A ma Cmaum	No. of Patients -	pН			
Age Group (yrs)	No. of Patients	Mean	Range 4-8 0 4-6·0		
10-20	36	5.7			
20-30	84	4.8			
30–40	62	5.2	4-7.0		
40-50+	18	5.2	46.5		

Table IV relates severity of symptoms to vaginal pH. It is clear that the women with the most severe symptoms and signs were those with the

Severity of Symptoms	No. of	pН	
Severity of Symptoms	Cases	Mean	Range
Asymptomatic, without vaginitis or discharge	56	5.9	4-8.0
Asymptomatic, with slight discharge but no vaginitis	34	5.7	4—7:5
Complaint of discharge, which is profuse but without vaginitis	64	4.8	45.5
Complaint of discharge and irritation; profuse discharge with definite vaginitis	48	4.4	4—5·5

Table IV
VAGINAL pH RELATED TO SEVERITY OF INFECTION

lowest vaginal pH. This does not appear to be due to their infections causing a lowering of vaginal pH (see Table II).

Cytologically, the patient with a low vaginal pH exhibits a vaginal aspirate rich in polymorphonuclear leucocytes, and a clustering of polymorphs around the epithelial cells may be observed. There may also be excessive maturation of the epithelial cells, and there is an impression of hyperchromasia. This condition may be due to increased vascularity (Koss and Durfee, 1961). Considerably more epithelial cells are found in those patients with a low vaginal pH. There may also be an increase in the size of the nuclei, but they retain their normal structure. It is of considerable importance to note that a low vaginal pH may exist in the relative absence or even total absence of Döderlein's bacilli, and it is suggested that the pH of the vagina depends on the degree of fermentation between the glycogen of the shed epithelial cells and the cervical mucus secretion. This would explain the increased morbidity in patients with a low vaginal pH in the presence of primary vaginal infections. The cytological picture of the vaginal exudate of the patient with a low vaginal pH corresponds to that found in women taking oral contraceptives, in diabetics, in pregnant women, and in those taking oral steroids. It seems reasonable to assume that, if excessive epithelium is being shed, invasion of the deeper layers may occur by normally benign saprophytic organisms which then will become "pathogenic".

It is suggested that any condition that increases the amount of cervical secretion or desquamation of vaginal epithelium may give rise to a primary vaginal discharge where the causative organism is able to penetrate through epithelial deficiencies. During this investigation it has been found that the woman with a low vaginal pH is in the first instance relatively resistant to treatment of a vaginal discharge; she is typically the patient who may require a second course of metronidazole for trichomoniasis or prolonged treatment for candidiasis. Conversely, the woman with a high vaginal pH may harbour yeasts or trichomonads and yet remain symptomless.

Summary

200 women, some with no genital disease, others with gonorrhoea, candidiasis, or trichomonal vaginitis, were studied by repeated measurements to determine possible variations in value of the vaginal pH. The principal finding was that each woman has a vaginal pH which remains stable within narrow limits and may only be altered, as far as present investigations show, by the administration of steroids. The pH does not appear to depend on the menarche, pregnancy, the menopause, illness, or the administration of other drugs. It is suggested that vaginal acidity depends on the amount of fermentation taking place between the glycogen-containing epithelial cells and the cervical mucus. It is possible that this amount of desquamation is regulated by the quantity of circulating oestrin. The woman with a low vaginal pH is more liable to increased morbidity from otherwise nonpathogenic saprophytic organisms and others causing primary vaginal discharges. This appears to be due to the excessive desquamation in those with a low vaginal pH causing a break in continuity of the epithelium, breaching the protective layers, and allowing penetration by the organisms.

REFERENCES

FEO, L. G. (1956). Amer. J. trop. Med. Hyg., 5, 786.

GALE, E. F. (1940). Bact. Rev., 4, 135.

_____, and Epps, H. M. R. (1942). Biochem. J., 36, 600.

Hunter, C. A., Jr., and Long, K. R. (1958). Amer. 3. Obstet. Gynec., 75, 872.

Koss, L. G., and Durfee, G. R. (1961). "Diagnostic Cytology and its Histopathologic Bases", p. 60. Lippincott, Philadelphia.

LUMSDEN, W. H. R., ROBERTSON, D. H. H., and Mc-NEILLAGE, G. J. C. (1966). Brit. J. vener. Dis., 42, 145.

RAKOFF, A. E., FEO, L. G., and GOLDSTEIN, L. (1944). Amer. J. Obstet. Gynec., 47, 467.

Stephenson, M. (1949). "Bacterial Metabolism", 3rd ed., pp. 97-98. Longmans Green, London.

Influence de pH sur les pertes vaginales

RÉSUMÉ

200 femmes, quelquesunes indemmes d'affection génitale, d'autres atteintes de gonococcie, de candidose ou de vaginite à trichomonas, furent l'objet de mesures répétées pour mettre en évidence des variations possibles de la valeur du pH vaginal. La principale constatation est que chaque femme a un pH vaginal qui reste stable dans d'étroites limites et qui peut seulement être changé, autant que le montre la présente recherche, par l'administration de stéroïdes. Le pH n'apparaît pas dépendre de la menstruation, de la grossesse, de la

ménopause, de maladies ou de l'administration d'autres médicaments. Il est suggéré que l'acidité vaginale dépend de l'importance de la fermentation qui se développe entre les cellules épithéliales chargées de glycogène et le mucus cervical. Il est possible que cette importance de la desquamation soit sous la dépendance de la quantité d'oestrine circulante.

Une femme à pH bas est plus exposée à la majoration de troubles morbides à partir de germes saprophytes non pathogènes ou d'autres responsables du début des pertes vaginales. Ceci semble être dû, chez celles à bas pH, à une desquamation excessive créant une solution de continuité de l'épithélium, rompant les couches protectrices et permettant la pénétration des germes.

APPENDIX

PROFORMA USED IN THE INVESTIGATION

NO A	.GE P	RESIDENTIA	L AREA	occui	PATION	SIBS	(Sisters	Brothers)
MARITAL STA	ATUS Marrie	d/Single/W	idowed/Divor	ced				
PREVIOUS DI	SCHARGES	AND TREA	ATMENT	• • • • • • • • • • • • • • • • • • • •		· · · · · · · · · · · · · · · ·		
SANITARY P	ROTECTION	External/Ir	nternal UN	NDERCLOT	HING Nyl	on/Cotton		
CONTRACEP	TIVE USED		• • • • • • • • • • • • • • • • • • • •					
MENARCHE.	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •	CYCLE		OI	BSTETRIC I	HISTORY	· · · · · · · · · · · · · · · · · · ·
DRUGS TAKE	N IN PAST	YEAR						
SEXUAL EXP	ERIENCE Ma	ırital/Extra	narital					
PAST HISTOR	RY General.	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •					• • • • • • • • • • • • • • • • • • • •
	Gynaecol	ogical	• • • • • • • • • • • • •				• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •
	Wet film	Film	Culture	рН	Tr	Date	Day of cycle	
								
						-		
				<u> </u>		<u>l</u>	<u> </u>	
WR PPR G	CFT OTHE	R SEROLO	GICAL TESTS	S •				
Cervical cytolo	ogy	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •	•••••	
Vulva	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •
Bartholin's gla	nds	• • • • • • • • • • • • • • • • • • • •				• • • • • • • • • • •		
Urethra				• • • • • • • • • • •				
Vagina								
Cervix				• • • • • • • • • • • • • • • • • • • •		• • • • • • • • • •		
Rectum and as	nus			• • • • • • • • • •				
Lymphatics				• • • • • • • • • •		• • • • • • • • • • •		
Extragenital le	esions			• • • • • • • • • •				
URINE SpGi	r. Alb. Su	gar Blood	Phosph. B	ile Aceton	e Alkaline	e/Acid Cas	sts Cells Dep	osits
TREATMENT	AND PROG	RESS	·			, 	·	
• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • •	• • • • • • • • • •	• • • • • • • • • • • •	• • • • • • • • • •	•••••	• • • • • • • • •	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • • • • • • • • • • • •